

# BRCA1/BRCA2: CanVIG-UK Gene-Specific Guidance



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For use in conjunction with CanVIG-UK Consensus Specification for Cancer susceptibility Genes of ACGS Best Practice Guidelines for Variant Classification. Evidence lines for which there are no gene-specific recommendations should be reviewed in context of CanVIG-UK Consensus Specification for Cancer Susceptibility Genes.

## Evidence towards Pathogenicity

Evidence element and evidence strengths allowed	Thresholds/data-sources/applications specifically relevant to BRCA1/BRCA2
<p><b>PS4: Case-control:</b> The prevalence of the variant in affected individuals is significantly increased compared with the prevalence in controls</p>	<p>NHSD case control data can be used for case-control analysis:</p> <ul style="list-style-type: none"> <li>• Controls should represent appropriate ethnicity and sex matching (i.e. female non-cancer NFE controls should be used if the case series consists predominantly of females, as with the current NHSD case series) and series denominator</li> <li>• As this is an enriched series, OR<math>\geq</math>10 is required</li> <li>• Current data/denominator counts for base substitutions are available at <a href="#">CanVar-UK</a></li> <li>• For non-base-substitutions i.e. deletions/duplications/insertions, NHSD counts can be accessed from <a href="#">CanVIG-UK</a></li> </ul> <p>If there are insufficient data to perform case-control analyses, PS4<sub>sup</sub> can be applied:</p> <ul style="list-style-type: none"> <li>• if there are observations of the variant in <math>\geq</math>5 different families with a pattern of diagnoses consistent with a hereditary breast and ovarian cancer syndrome</li> <li>• <b>and</b> the variant is very rare or absent in control populations (i.e. PM2 has been applied)</li> </ul>
<p><b>PM2: Absent from controls</b> (or at extremely low frequency if recessive) in ESP, 1000GP, or ExAC</p>	<p>Cancer-free female controls (of any/all ethnicities) should be used (due to low penetrance in male pathogenic variant carriers). Otherwise, the main CanVIG-UK consensus guidance should be followed.</p>

<p><b>PVS1: Predicted null variant</b> (in a gene where LOF is a known mechanism of disease)</p>	<p>_VSTR _STR _MOD _SUP</p>	<p>It is predicted that truncating variants occurring at the 3' end of the gene will not undergo NMD. The residues below demarcate the consensus boundary, 3' of which protein truncating variants are not established to result in NMD and/or impairment of function of residual protein. BRCA1 (NM_007294.3): 1855<sup>1</sup> BRCA2 (NM_000059.3): 3309<sup>2</sup> Based on ENIGMA recommendations, as re-initiation sites have also been shown to result in the loss of important functional domains in BRCA1 and BRCA2, it is acceptable to use PVS1 at a very strong level for variants identified within the first 100bp of both BRCA1 and BRCA2<sup>1</sup>. A number of variants at canonical splice sites are predicted or known to lead to naturally occurring in-frame RNA isoforms that may rescue gene functionality. ENIGMA has compiled the below list of splice variants for which the variant transcript may be functional and for some of which PVS1 may not be applicable.</p> <table border="1" data-bbox="643 680 1042 1310"> <thead> <tr> <th>Gene</th> <th>Region</th> <th>Bases</th> </tr> </thead> <tbody> <tr> <td rowspan="12">BRCA1</td> <td>intron 5 (exon 5 donor)</td> <td>c.301+1 c.301+2</td> </tr> <tr> <td>intron6 (exon 7 acceptor)</td> <td>c.442-1 c.442-2</td> </tr> <tr> <td rowspan="6">introns 8,9</td> <td></td> <td>c.548-1 c.548-2 c.593+1 c.593+2 c.594-1 c.594-2</td> </tr> <tr> <td></td> <td>c.670+1 c.670+2</td> </tr> <tr> <td>intron 10 (exon 10 donor)</td> <td>c.4096+1 c.4096+2</td> </tr> <tr> <td>intron 11 (exon 12 acceptor)</td> <td>c.4186-1 c.4186-2</td> </tr> <tr> <td>intron12 (exon 13 acceptor)</td> <td>c.4358-1 c.4358-2</td> </tr> <tr> <td rowspan="4">BRCA2</td> <td rowspan="4">intron12</td> <td>c.6842-1 c.6842-2 c.6937+1 c.6937+2</td> </tr> </tbody> </table> <p>Adapted from ENIGMA, 2017<sup>1</sup></p>	Gene	Region	Bases	BRCA1	intron 5 (exon 5 donor)	c.301+1 c.301+2	intron6 (exon 7 acceptor)	c.442-1 c.442-2	introns 8,9		c.548-1 c.548-2 c.593+1 c.593+2 c.594-1 c.594-2		c.670+1 c.670+2	intron 10 (exon 10 donor)	c.4096+1 c.4096+2	intron 11 (exon 12 acceptor)	c.4186-1 c.4186-2	intron12 (exon 13 acceptor)	c.4358-1 c.4358-2	BRCA2	intron12	c.6842-1 c.6842-2 c.6937+1 c.6937+2
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	<p><b>PS1: Same amino acid change</b> as an established variant</p>			<p>_STR</p>	<p>Within forthcoming ENIGMA guidance it is anticipated that these elements will all be incorporated within PP3 and only awarded to variants within key domains:</p>																			
	<p><b>PM4: Protein-length-changing variant</b></p>			<p>_MOD _SUP</p>	<p><b>In the interim, we recommend:</b></p> <ul style="list-style-type: none"> <li>Use of PM1_sup and/or PM4_sup for any variant within BRCA1 RING (αα 1-101), BRCT (αα1650-1863) COILED-COIL DOMAIN (αα 1391-1424) and BRCA2 DNA-binding domain (αα 2481-3186)</li> </ul>																			
	<p><b>PM5: Novel missense change</b> at an amino acid residue where a different missense change determined to be pathogenic seen before</p>			<p>_MOD _SUP</p>	<ul style="list-style-type: none"> <li>Use of PM1_mod for missense at specific residues<sup>3</sup>: RING: 18, 22, 37, 39, 41, 44, 47, 61, 64, 71 BRCT: 1685, 1688, 1697, 1699, 1706, 1708, 1715, 1736, 1738, 1739, 1748, 1764, 1766, 1770, 1775, 1786, 1837, 1838, 1839, 1853 DBD: 2607, 2626, 2627, 2663, 2722, 2723, 2748, 3052, 3124</li> </ul>																			
	<p><b>PP3: In silico:</b> Multiple lines of computational evidence support a deleterious effect on the gene or gene product</p>	<p>_SUP</p>																						
<p><b>PM1, PP2: Enrichment/constraint:</b> <b>PP2:</b> Missense variant in a gene that has a low rate of benign missense variation and in which missense variants are a common mechanism of disease</p>	<p>_STR _MOD _SUP</p>	<ul style="list-style-type: none"> <li>PM1 cannot be used where functional data are being used for PS3, as per main CanVIG-UK guidance</li> <li>PP2 should not be used for BRCA1/BRCA2</li> <li>Use of PM5, PS1, PP3 otherwise as per CanVIG-UK Consensus Specification</li> </ul>																						

<p><b>PM1:</b> Located in a mutational hot spot and/or critical and well-established functional domain (e.g. active site of an enzyme) without benign variation</p>		
<p><b>PS3: Functional:</b> Well-established in vitro or in vivo functional studies supportive of a damaging effect on the gene or gene product</p>	<p><b>_VSTR</b> <b>_STR</b> <b>_MOD</b> <b>_SUP</b></p>	<p><b>BRCA1:</b> Findlay et al, 2018<sup>4</sup>: Strong Bouwman et al, 2020<sup>5</sup>: Strong Fernandes et al, 2019<sup>6</sup>: Supporting Petitalot et al, 2019<sup>7</sup>: Supporting <b>BRCA2:</b> Guidugli et al, 2018<sup>8</sup>/Hart et al, 2019<sup>9</sup>/Richardson et al, 2021<sup>10</sup>: Strong <a href="#">See CanVIG Functional Assays Scores</a> See the table at the bottom of this document for guidance on combining assay results</p>
<p><b>PP1: Co-segregation</b> with disease in multiple affected family members in a gene definitively known to cause the disease</p>	<p><b>_VSTR</b> <b>_STR</b> <b>_MOD</b> <b>_SUP</b></p>	<p>Segregation evidence extracted from multifactorial analysis data can be used within PP1/BS4 using the thresholds specified in the PP5/BP6 guidance. Where combined with multiple evidence of other types, segregation evidence from multifactorial analysis data should be incorporated into the PP5/BP6 criteria Meiosis counting approaches may be used in addition if this evidence comes from families not already included in the multifactorial analyses. <b>Evidence cannot exceed ‘Very strong’</b></p>
<p><b>PS2/PM6: De novo</b> (maternity and paternity confirmed/unconfirmed) in a patient with the disease and no family history</p>	<p><b>_STR</b> <b>_MOD</b> <b>_SUP</b></p>	
<p><b>PM3: in trans</b> with a pathogenic variant (<b>recessive disorders</b>)</p>	<p><b>_STR</b> <b>_MOD</b> <b>_SUP</b></p>	<p>Frequency data regarding co-occurrence in trans extracted from multifactorial analyses should be incorporated into PM3 or BP2 using the thresholds described in the PP5/BP6 guidance. Where combined with multiple evidence of other types, frequency data regarding co-occurrence in trans from multifactorial analyses should be incorporated into PP5/BP6</p> <p><b>In addition</b>, the <a href="#">SVI recommendations for in trans Criterion (PM3)</a> can be used for either BRCA1 or BRCA2 for individuals with a Fanconi anaemia phenotype if this evidence comes from families not already included in the multifactorial analyses used for PP5. Evidence towards a Fanconi phenotype comprise:</p> <ul style="list-style-type: none"> <li>• <b>Clinical:</b> diagnosis of childhood cancer or skeletal/structural/developmental abnormalities</li> <li>• <b>Molecular/Cellular:</b> aberration on mitomycin-induced chromosomal breakage +/- depletion of BRCA2 in lymphocytes</li> </ul> <p>Both clinical and molecular/cellular aberrations must be present for a case to contribute to evidence <b>Evidence cannot exceed ‘Strong’</b></p> <p><b>Note:</b> Caution is required in inferring the pathogenicity for the monoallelic phenotype, as variants may be hypomorphic (e.g. a variant contributing and causing a Fanconi anaemia phenotype may be low penetrance for breast cancer). Where the majority of evidence for variant pathogenicity comes from observations of the variant in cases of Fanconi Anaemia, it may be appropriate to comment on this in the clinical report</p>

**PP5: Reputable source**  
recently reports variant as pathogenic, but the evidence is not available to the laboratory to perform an independent evaluation

\_VSTR  
\_STR  
\_MOD  
\_SUP

Published multifactorial analysis data providing likelihood ratios (LR) or log likelihood ratios (LLR) may be used as data sources encompassing:

- Segregation (PP1/BS4)
- Specificity of familial and/or tumour phenotype (PP4)
- Co-occurrence in trans (PM3/BP2)

Where individual likelihood ratios for a particular evidence type do not line up with evidence (exponent) points required for a specific evidence strength, the **combined LLR/LR** encompassing multiple evidence types can be used instead to represent the totality of evidence and applied within PP5

Suitable analyses:

- Easton et al, 2007<sup>11</sup>
- Parsons et al, 2020<sup>12</sup>

Where multiple potentially valid LR/LLRs are available for a variant, the value from the most recent publication should be used.

Where evidence is supplied as a LR (likelihood ratio, e.g. Parsons et al, 2020) this should be converted to Evidence (Exponent) points using the table below.

Where evidence is supplied as a natural LLR (log likelihood ratio, e.g. Easton et al, 2007), this should be converted to a LR (for example using the =EXP() function in excel) before conversion to Evidence (Exponent) Points using the table below (i.e. converted from the LR to a LLR base 2.08)

Likelihood Ratio	Evidence (Exponent) Points	Evidence Strength
2.1	1	SUP
4.3	2	MOD
9	3	
18.7	4	STR
38.9	5	
81	6	
168.4	7	
350.4	8	VSTR

**OR**

PP5 can be applied at supporting level on the basis of any classification of LP/P after 2018 using ACMG classification from:

- ≥2 accredited North American commercial diagnostic laboratories OR
- ≥1 North American commercial diagnostic laboratory where there is explicit citation of utilisation of otherwise unavailable evidence from their data series OR
- approved ClinGen Expert Group (3 star on ClinVar), ie ENIGMA

This is an **exceptional** application, as per UK-ACGS specification.

For conflicts with ENIGMA classifications, contact ENIGMA.

**PP4: Phenotypic specificity** (Patient's

\_STR  
\_MOD

Tumour and family history phenotypic data extracted from multifactorial analyses should be incorporated into PP4 or BP5

phenotype or family history is highly specific for a disease with a single genetic aetiology)	_SUP	using the thresholds described in the PP5/BP6 guidance. Where combined with multiple evidence of other types, tumour and family history phenotypic data from multifactorial analyses are incorporated into the PP5 evidence criterion. Patient phenotypic evidence whose strength cannot be quantified should not be used. <b>Evidence cannot exceed ‘Strong’</b>
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### Evidence towards Benignity

<b>BA1/BS1: Allele frequency</b> is “too high” in ExAC or gnomAD for disorder	_SA _STR	<b>BA1: MTAf = 0.001 (0.1%)</b> <b>BS1: MTAf = 0.0001 (0.01%)</b> The U95%CI should be used as the filtering allele count for the MTAf. This can be calculated using <a href="#">cardiodb</a> or within gnomAD (see <a href="#">training resources</a> from Miranda Durkie for methodology) Cancer-free <b>female controls</b> should be used (due to low penetrance in male pathogenic variant carriers)
<b>BS2: Observation in controls</b> inconsistent with disease penetrance. Observed in a healthy adult individual for a recessive (homozygous), dominant (heterozygous), or X-linked (hemizygous) disorder, with full penetrance expected at an early age	_STR _SUP	
<b>BP4: In silico:</b> Multiple lines of computational evidence suggest no impact on gene or gene product (conservation, evolutionary, splicing impact, etc.)	_SUP	
<b>BP1: Missense variant in a gene for which primarily truncating variants are known to cause disease</b>	_SUP	Can be used for missense variants with no predicted splicing effect (as per main CanVIG-UK consensus specification) at non-conserved residues outside of BRCA1 RING (aa 1-101), BRCT (aa 1650-1863) COILED-COIL DOMAIN (aa 1391-1424) and BRCA2 DNA-binding domain (aa 2481-3186)
<b>BP7: Synonymous</b> (silent) variant for which splicing prediction algorithms predict no impact to the splice consensus sequence	_SUP	
<b>BP3: In-frame deletions/insertions in a repetitive region</b>	_SUP	
<b>BS3: Well-established <i>in vitro</i> or <i>in vivo</i> functional studies</b> show no damaging effect on protein function or splicing	_STR _MOD _SUP	
<b>BS4: Non segregation with disease</b>	_STR _SUP	*see PP1
<b>BP2: Observed in trans with a pathogenic variant</b> for a fully penetrant dominant gene/disorder or observed in cis	_STR _SUP	*see PM3
<b>BP6: Reputable source</b> recently reports variant as	_STR _SUP	*see PP5

benign, but the evidence is not available to the laboratory to perform an independent evaluation			
		Likelihood Ratio	Evidence (Exponent) Points
		0.48	-1
		0.23	-2
		0.11	-3
		0.05	-4
			Evidence Strength
			SUP
			STR
<b>BP5:</b> Alternate molecular basis for disease	<u>_SUP</u>	*see PP4	

**Recommendations for the management of conflicting functional assay results** See table below for management of discrepancy for BRCA1 variants between Findlay et al, 2018<sup>4</sup> and Bouwman et al, 2020<sup>5</sup> discordant assay results. For more general guidance regarding conflicting results from other functional assays, refer to the table in the main CanVIG-UK consensus specification.

Findlay Class	Findlay Score	Bouwman Platinum	Bouwman Olaparib	Bouwman DR-GFP	PS3_STR	BS3_STR
LOF	<-1.328	All deleterious/ likely deleterious (1 intermediate allowed)			✓	✗
LOF	<-1.328	Any are neutral/likely neutral			✗	✗
INT (towards LOF)	-1.328 to -1.038	All deleterious/ likely deleterious			✓	✗
INT (towards FUNC)	-1.038 to -0.748	All neutral/likely neutral			✗	✓
INT	-1.328 to -0.78	Conflicting results or any intermediate			✗	✗
FUNC	>-0.748	All neutral/likely neutral (1 intermediate allowed)			✗	✓
FUNC	>-0.748	Any are deleterious/likely deleterious			✗	✗

N.B: Bouwman et al, 2020 “not clear” refers to opposite categorisation ± the standard deviation of repeat experiments and should be treated as conflicting assay results. Where a variant is LOF on the Findlay et al assay and has an RNA score of <-2, this indicates that LOF is due to interference with splicing and therefore should not be treated as conflicting evidence if the variant is neutral on the Bouwman et al assay.

### Version History/Amendments

Revised version	Date	Section	Update	Amended by	Approved by
1.12	01/09/2021	PP4	Guidance on use of LLRs from published epidemiological studies amended to account for the use of natural logs in the statistics presented	Garrett	Turnbull
1.12	01/09/2021	PM1	Addition of critical residues in the DNA binding domain of BRCA2. Critical residues in all listed functional domains updated to mirror draft 2021 ENIGMA guidance	Garrett	Turnbull
1.12	01/09/2021	BP1	Resolution of typo in BRCT region specification	Garrett	Turnbull
1.13	15/10/2021	PVS1	Clarification that PVS1 may not be applicable for some of the variants at ENIGMA specified positions	Garrett	Turnbull
1.14	02/12/2021	PS4	Terminology change to reflect transition of PHE to NHSD	Garrett	Turnbull



1.14	02/12/2021	PVS1	Addition of recommendations for variants within the first 100bp	Callaway	CStAG
1.15	28/04/2022	PM1/PM4	Clarification that PM1_sup and PM4_sup may be used in combination but PM4 not to be used at moderate. Removal of mention that CanVIG-UK provide non-white ethnicity counts under PS4	Garrett	CStAG
1.16	28/07/2022	PP5	Removal of Lindor <i>et al</i> 2011 paper from recommended genetic epidemiology papers to use in calculating Evidence (Exponent) Points.	Allen	Turnbull

## References

1. Draft ACMG/AMP Classification Rules Specified for BRCA1 & BRCA2 ENIGMA Variant Curation Expert Panel, Classification Criteria V1.0 2021-06-21., 2021.
2. Mesman RLS, Calléja F, Hendriks G, et al. The functional impact of variants of uncertain significance in BRCA2. *Genetics in medicine : official journal of the American College of Medical Genetics* 2019;21(2):293-302. doi: 10.1038/s41436-018-0052-2 [published Online First: 2018/07/11]
3. ENIGMA. BRCA1/2 Gene Variant Classification Criteria Version 2.5.1 2017 [Available from: <https://enigmaconsortium.org/library/general-documents/enigma-classification-criteria/>].
4. Findlay GM, Daza RM, Martin B, et al. Accurate classification of BRCA1 variants with saturation genome editing <https://sge.gs.washington.edu/BRCA1/>. *Nature* 2018;562(7726):217-22. doi: 10.1038/s41586-018-0461-z [published Online First: 2018/09/14]
5. Bouwman P, van der Heijden I, van der Gulden H, et al. Functional Categorization of BRCA1 Variants of Uncertain Clinical Significance in Homologous Recombination Repair Complementation Assays. *Clinical cancer research : an official journal of the American Association for Cancer Research* 2020;26(17):4559-68. doi: 10.1158/1078-0432.Ccr-20-0255 [published Online First: 2020/06/18]
6. Fernandes VC, Golubeva VA, Di Pietro G, et al. Impact of amino acid substitutions at secondary structures in the BRCT domains of the tumor suppressor BRCA1: Implications for clinical annotation. *The Journal of biological chemistry* 2019;294(15):5980-92. doi: 10.1074/jbc.RA118.005274 [published Online First: 2019/02/16]
7. Petitalot A, Dardillac E, Jacquet E, et al. Combining Homologous Recombination and Phosphopeptide-binding Data to Predict the Impact of BRCA1 BRCT Variants on Cancer Risk. *Mol Cancer Res* 2019;17(1):54-69. doi: 10.1158/1541-7786.Mcr-17-0357 [published Online First: 2018/09/28]
8. Guidugli L, Shimelis H, Masica DL, et al. Assessment of the Clinical Relevance of BRCA2 Missense Variants by Functional and Computational Approaches. *American journal of human genetics* 2018;102(2):233-48. doi: 10.1016/j.ajhg.2017.12.013 [published Online First: 2018/02/06]
9. Hart SN, Hoskin T, Shimelis H, et al. Comprehensive annotation of BRCA1 and BRCA2 missense variants by functionally validated sequence-based computational prediction models. *Genetics in medicine : official journal of the American College of Medical Genetics* 2019;21(1):71-80. doi: 10.1038/s41436-018-0018-4 [published Online First: 2018/06/10]
10. Richardson ME, Hu C, Lee KY, et al. Strong functional data for pathogenicity or neutrality classify BRCA2 DNA-binding-domain variants of uncertain significance. *American journal of human genetics* 2021;108(3):458-68. doi: 10.1016/j.ajhg.2021.02.005 [published Online First: 2021/02/21]
11. Easton DF, Deffenbaugh AM, Pruss D, et al. A systematic genetic assessment of 1,433 sequence variants of unknown clinical significance in the BRCA1 and BRCA2 breast cancer-predisposition genes. *American journal of human genetics* 2007;81(5):873-83. doi: 10.1086/521032 [published Online First: 2007/10/10]
12. Parsons MT, Tudini E, Li H, et al. Large scale multifactorial likelihood quantitative analysis of BRCA1 and BRCA2 variants: An ENIGMA resource to support clinical variant classification. *Human mutation* 2019;40(9):1557-78. doi: 10.1002/humu.23818 [published Online First: 2019/05/28]